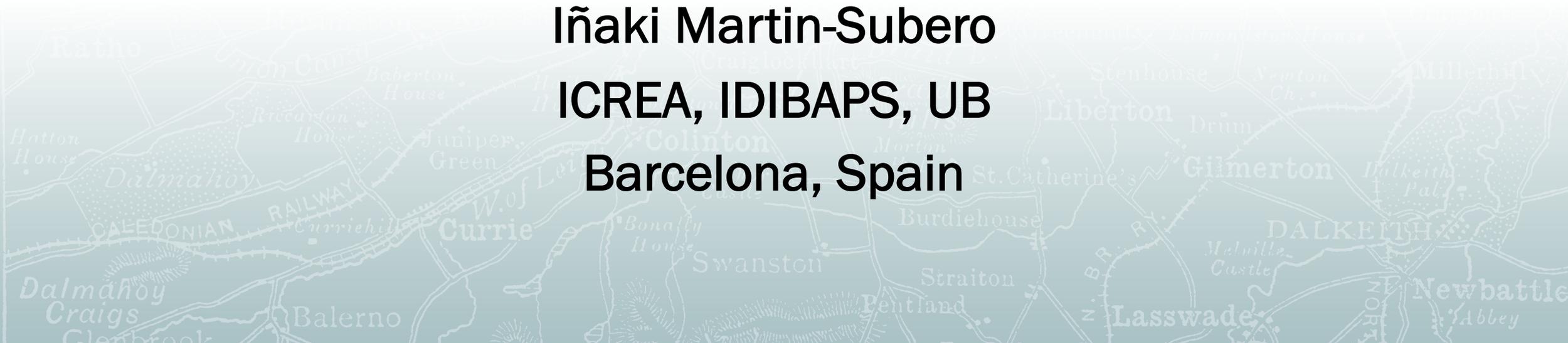




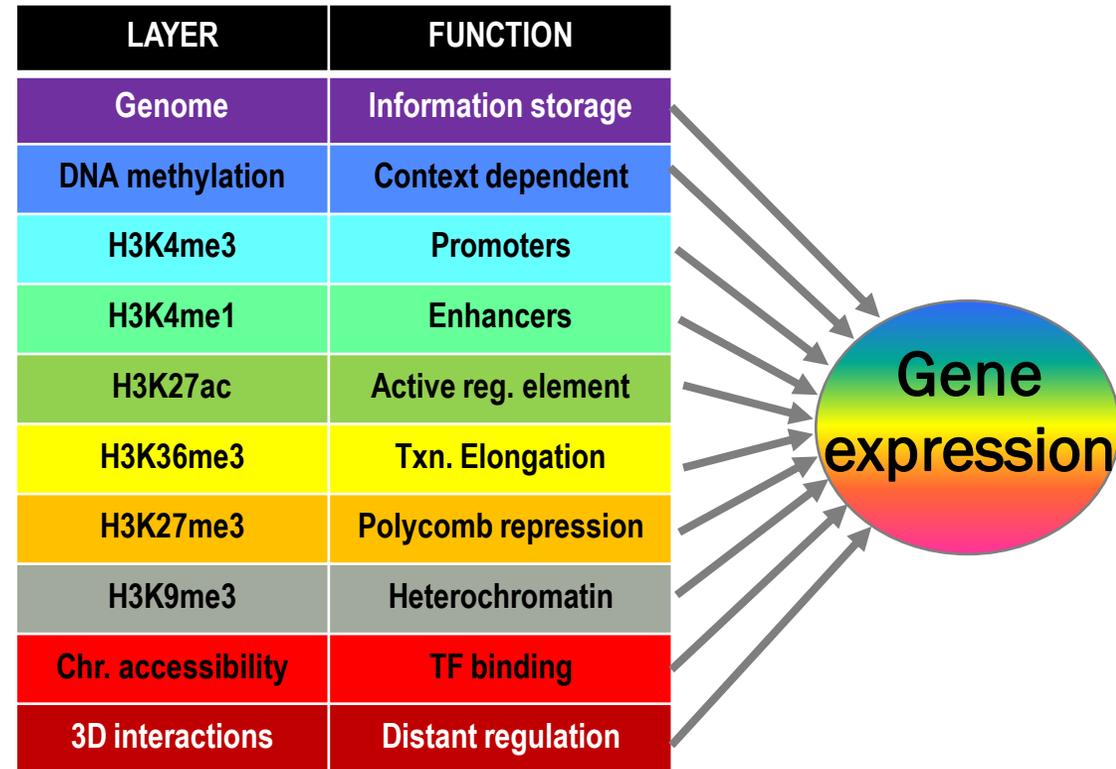
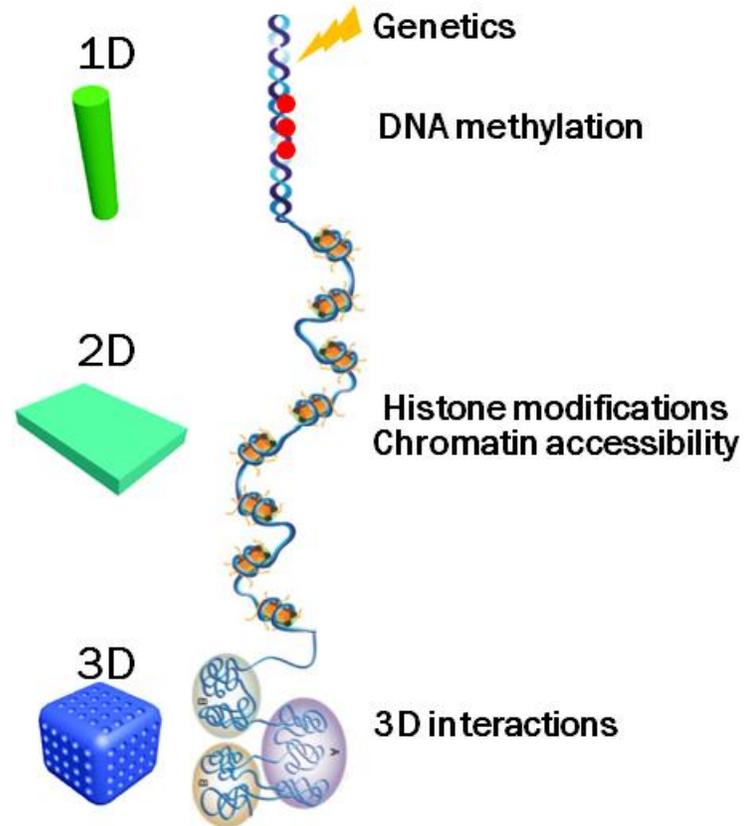
International Workshop on CLL  
20-23 SEPTEMBER 2019 EDINBURGH

# *Insights from Tertiary Structure, Chromatin Modifications, and Nuclear Organization*

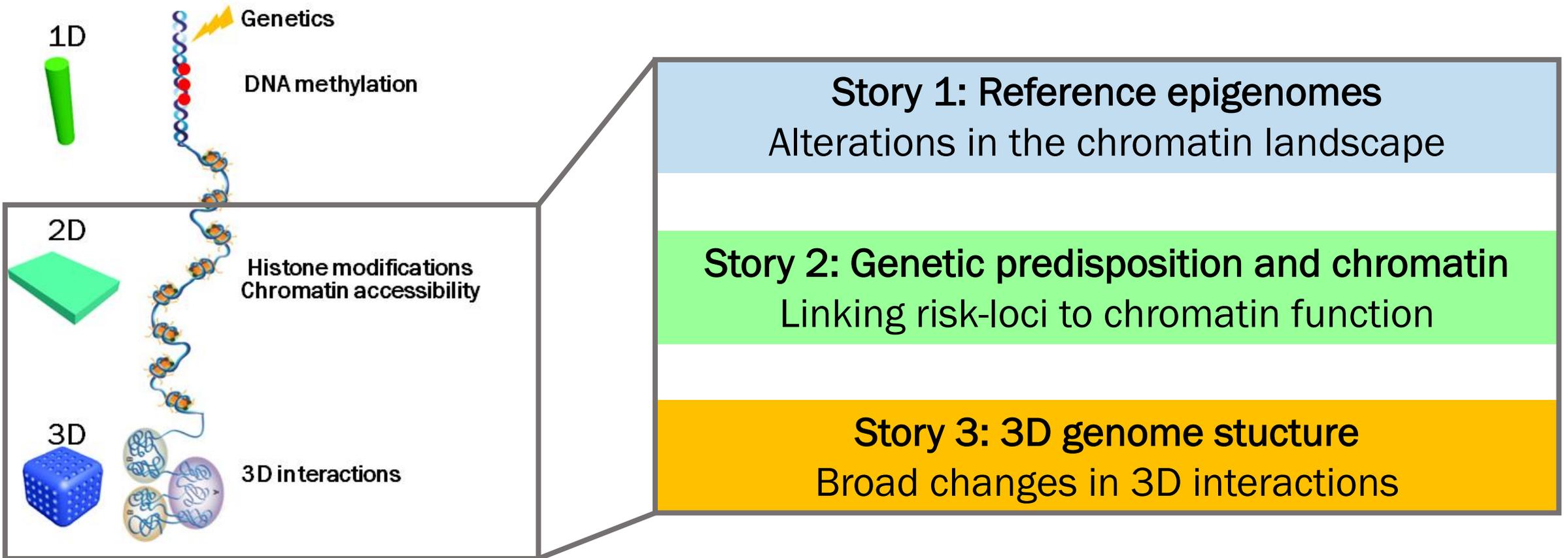
**Iñaki Martin-Subero**  
**ICREA, IDIBAPS, UB**  
**Barcelona, Spain**



# Epigenetic information controls gene expression

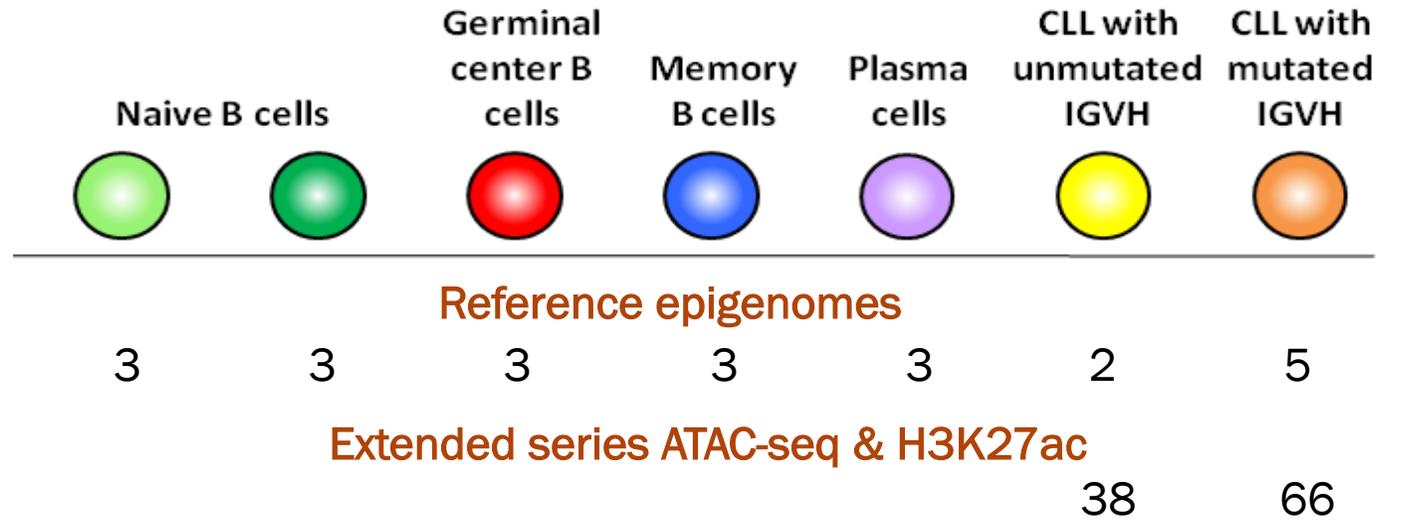
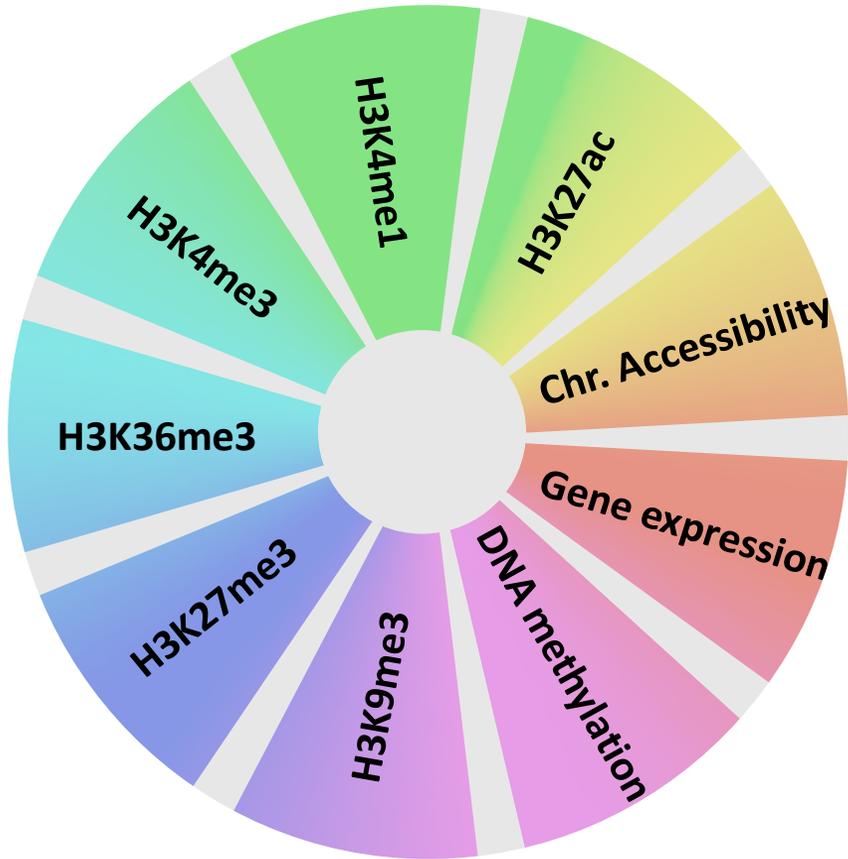


# Three epi & genetic stories

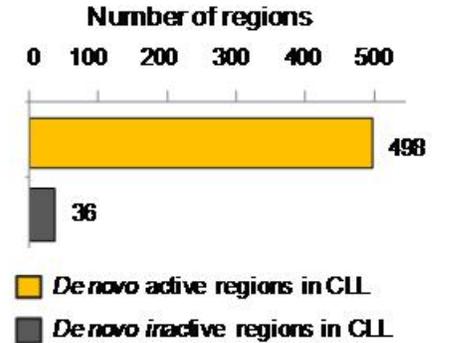
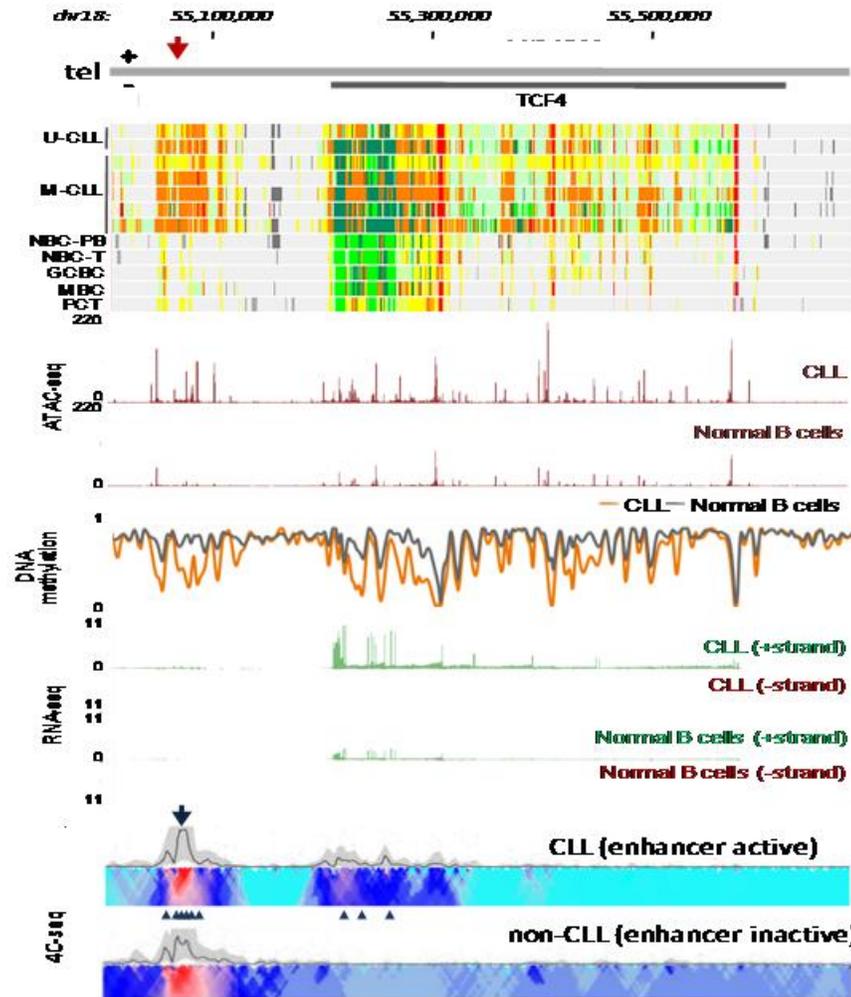
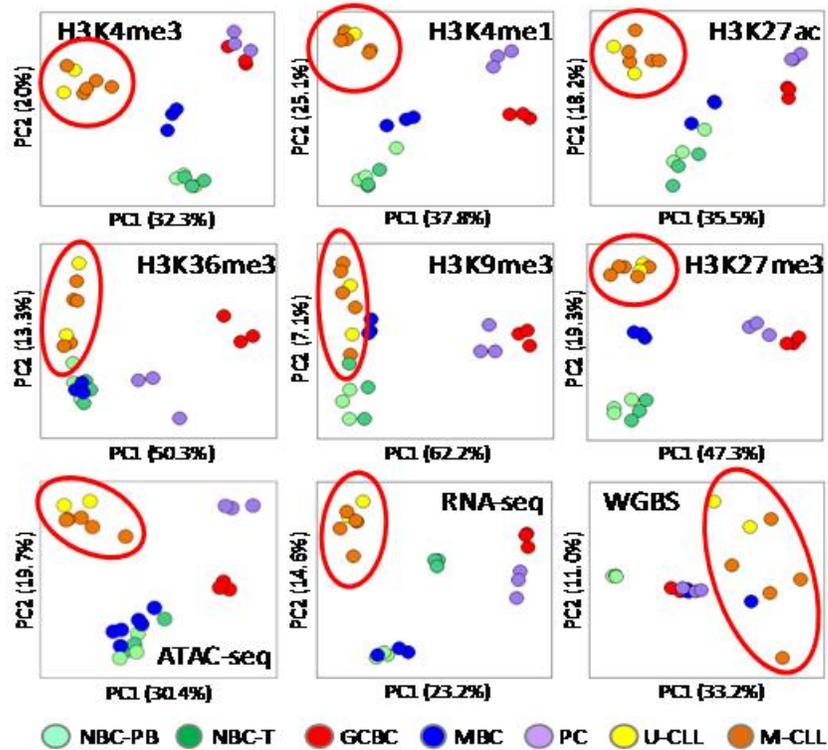




# Story 1: Reference epigenomes of CLL



# Story 1: De novo chromatin activation in CLL



NFAT family (P val=6e-16)



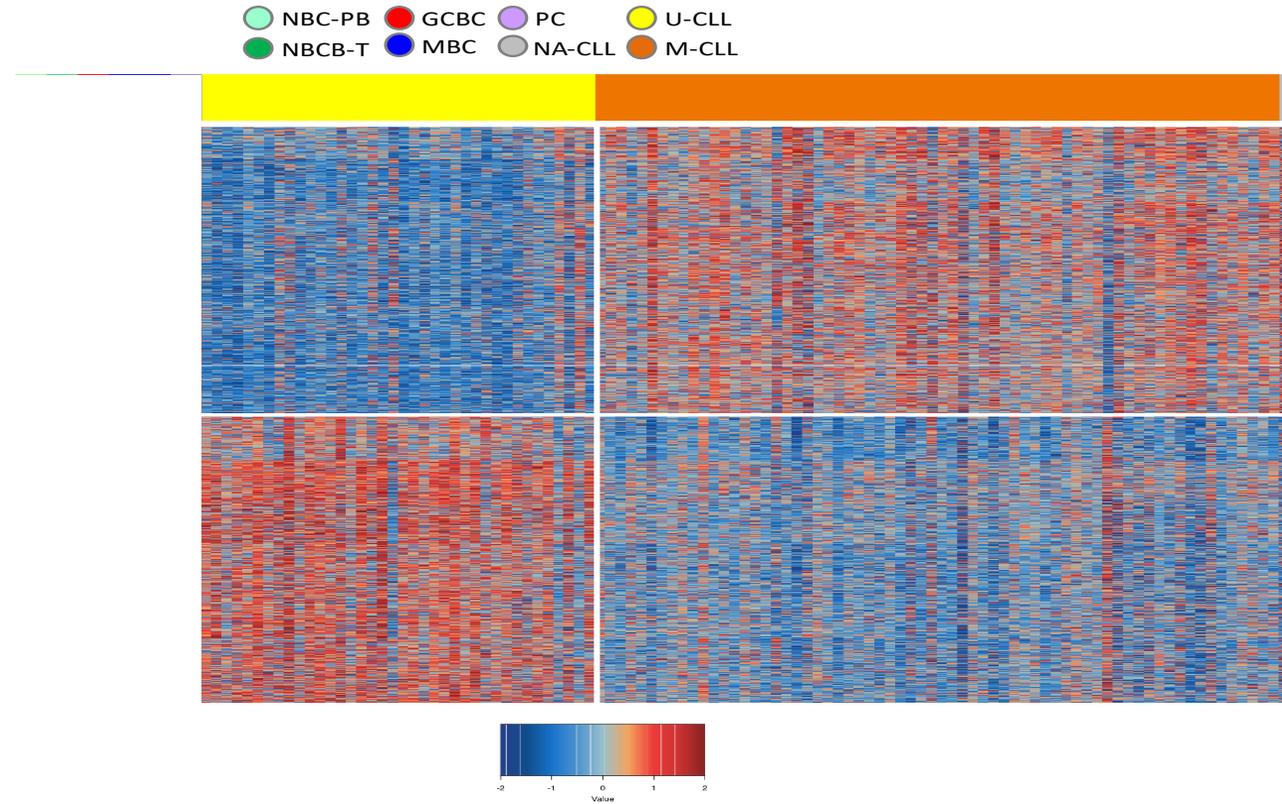
FOX family (P val=1e-11)



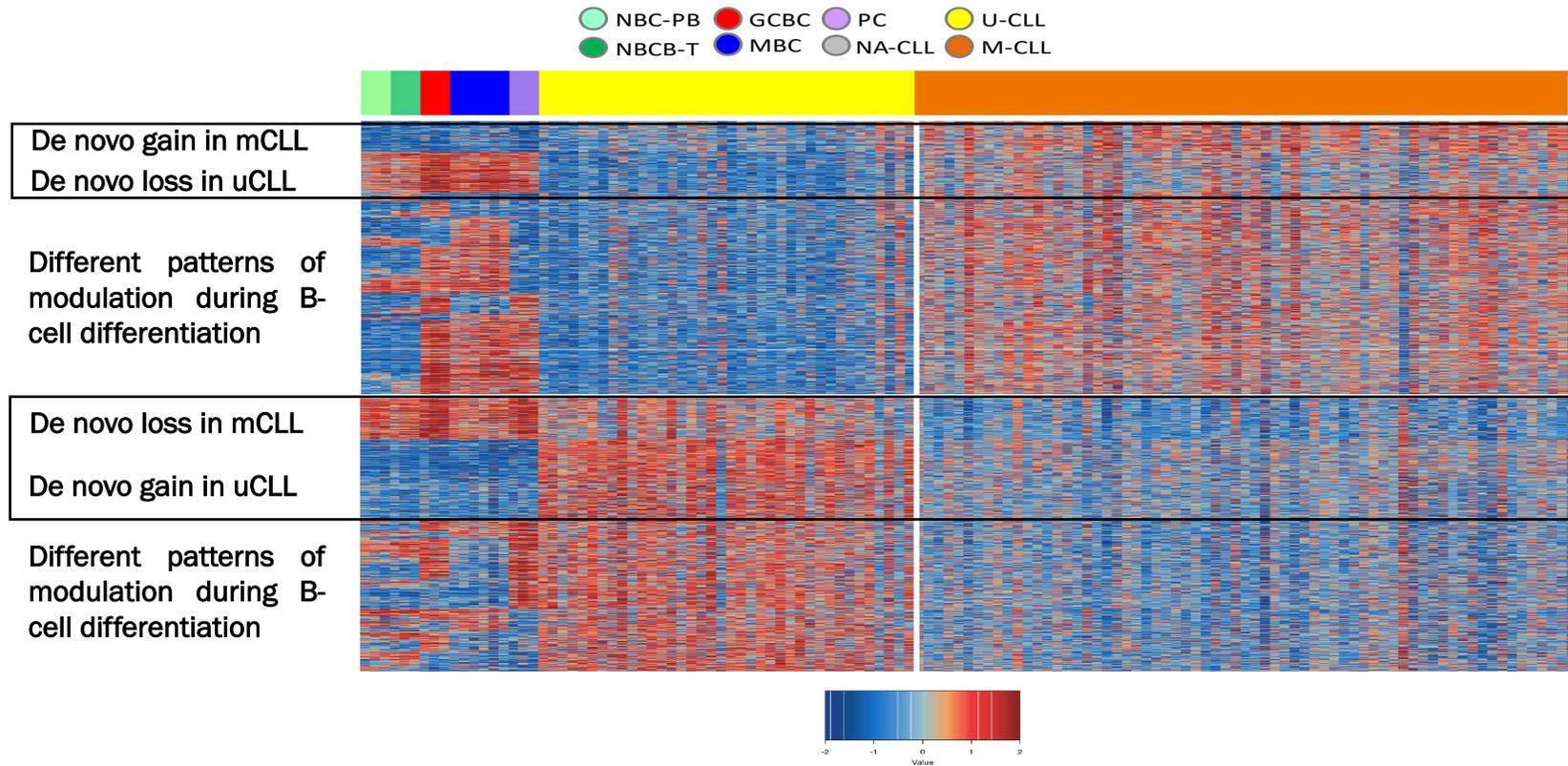
TCF/LEF family (P val=5e-9)



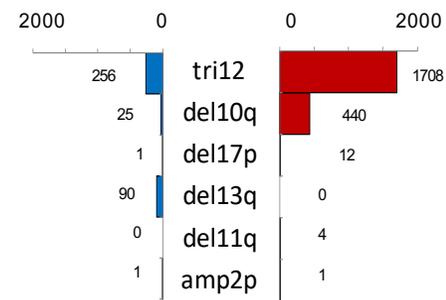
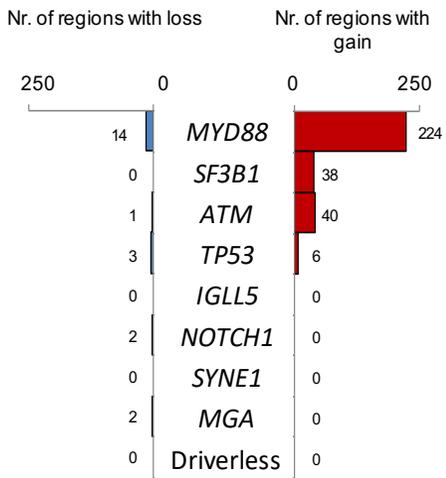
# Story 1: Different chromatin landscapes in uCLL and mCLL



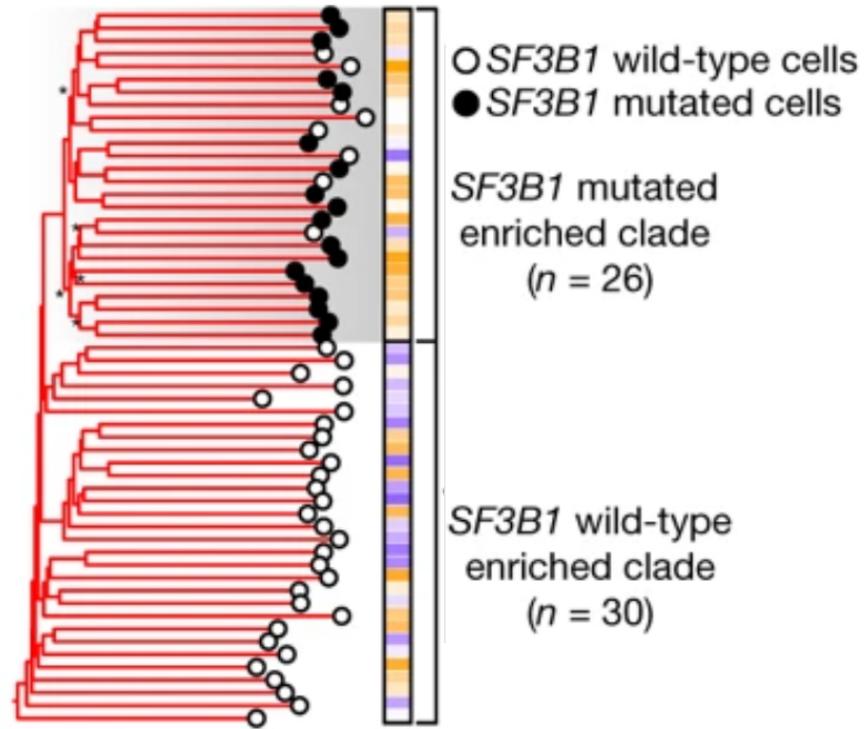
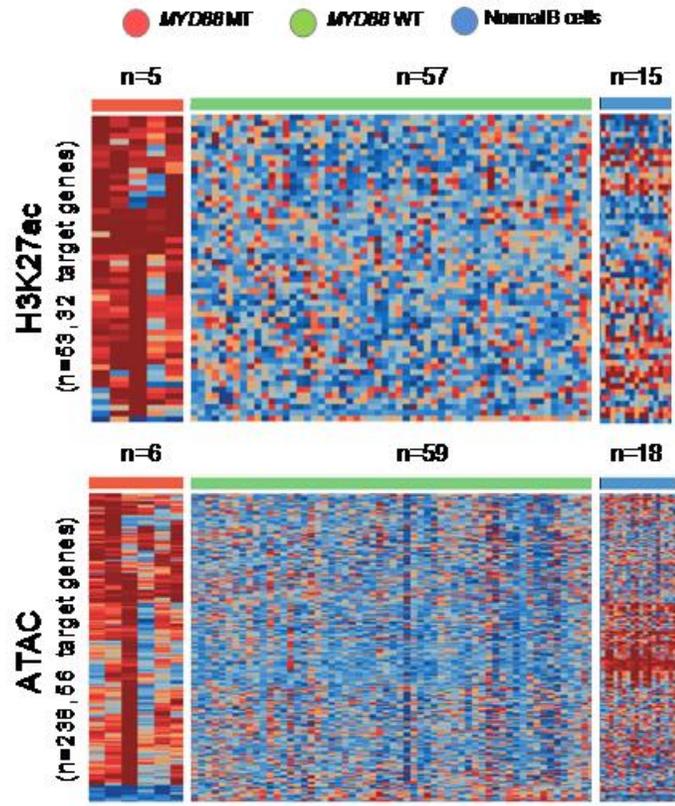
# Story 1: Different chromatin landscapes in uCLL and mCLL



# Story 1: Association between genetic changes and chromatin



ATAC



Gaiti et al., Nature 2019

Beekman et al., Nat Med 2018

# Story 2: Genetic predisposition and chromatin

**The reference epigenome and regulatory chromatin landscape of chronic lymphocytic leukemia**

Renée Beekman<sup>1,2</sup>, Vicente Chapaprieta<sup>3</sup>, Núria Russiñol<sup>1</sup>, Roser Vilarrasa-Blasi<sup>3</sup>, et al.

Genome-wide association analysis implicates dysregulation of immunity genes in chronic lymphocytic leukaemia

Phillip J. Law et al.<sup>#</sup>

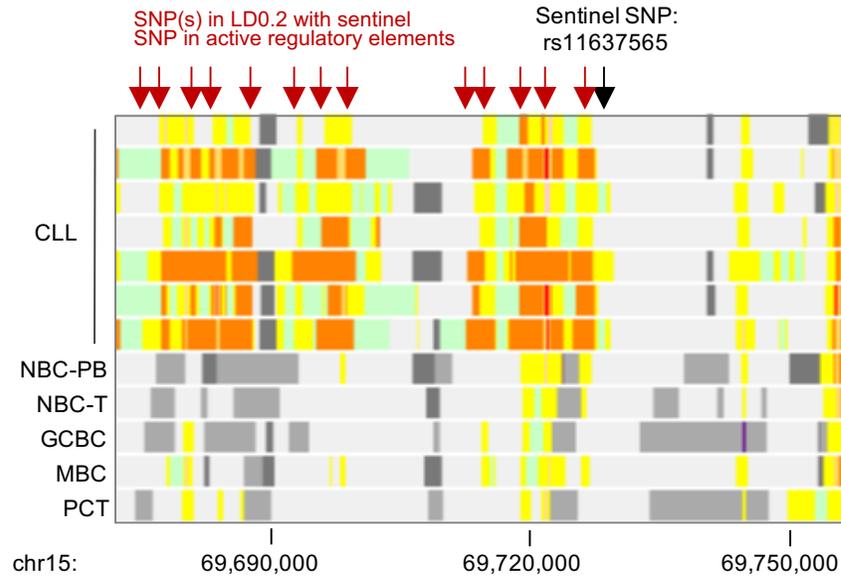
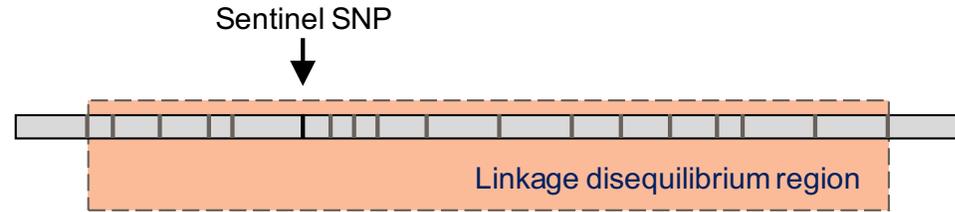
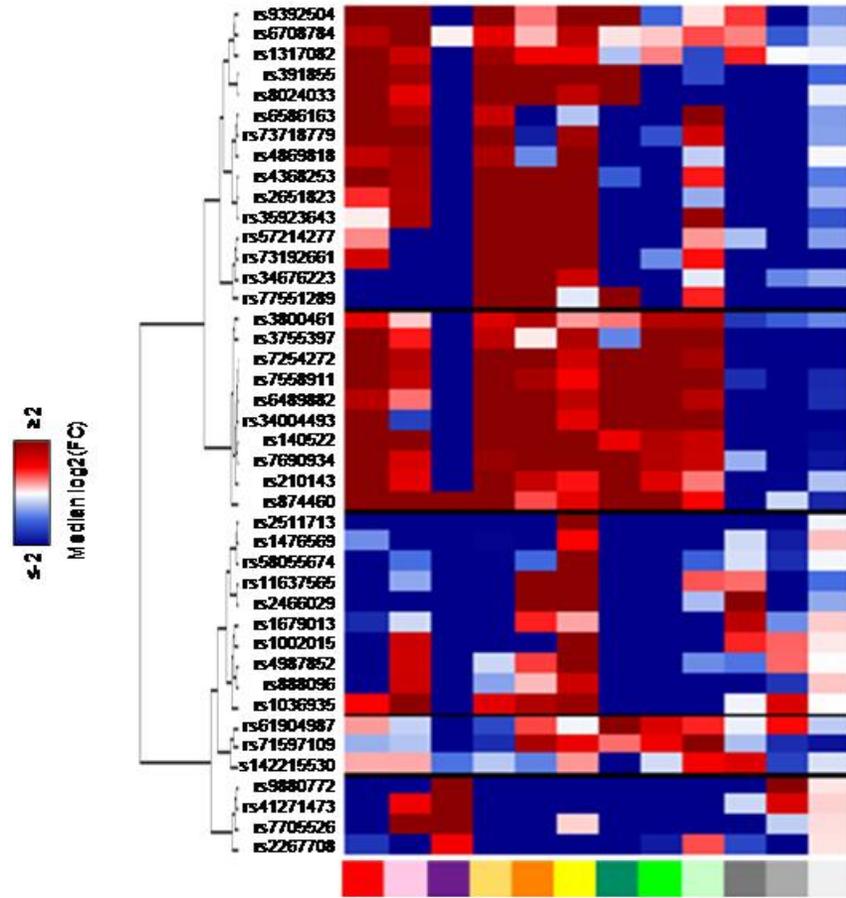


Richard Houlston

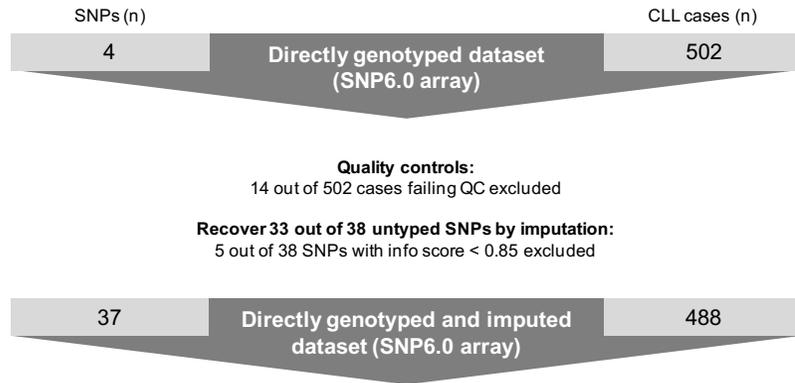
Can the chromatin landscape help us to better understand genetic predisposition?



# Story 2: Most risk-loci are located in regulatory elements

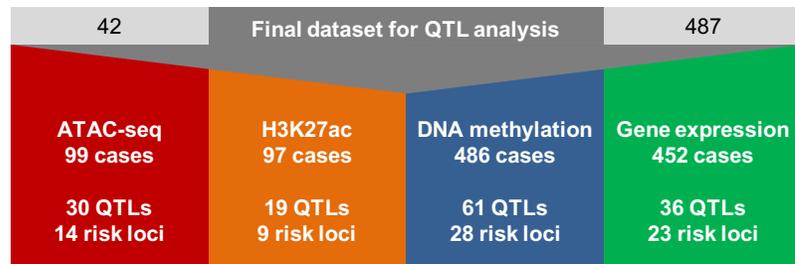


# Story 2: Target genes & altered TF binding affinity



Use WGS data of 146 out of 488 cases to check fidelity of imputation:  
1 case with discordance > 10% excluded, >99% concordance across all other genotypes

Complete missing genotypes for remaining 145 out of 487 cases with WGS data:  
Including genotypes of 5 SNPs with imputation info score < 0.85



QTL analysis

SNP	Position	ATAC-seq	H3K27ac	DNA methylation	Gene expression	Candidate target genes
rs35923643	chr11:123484683	■	■	■	■	GRAMD1B
rs6489882	chr12:112943571	■	■	■	■	OAS1, OAS3
rs11637565	chr15:69728186	■	■	■	■	TLE3
rs2511713	chr8:102565637	■	■	■	■	UBR5
rs7558911	chr2:201159226	■	■	■	■	CASP8, CFLAR, PP1L3
rs7690934	chr4:108104709	■	■	■	■	LEF1
rs1679013	chr9:22206988	■	■	■	■	DMRTA1
rs8024033	chr15:40111456	■	■	■	■	BMF
rs391855	chr16:85895015	■	■	■	■	IRF8
rs3755397	chr2:241355498	■	■	■	■	SEPT2, ING5, MTERF4
rs140522	chr22:50532837	■	■	■	■	CHKB-CPT1B, CPT1B, ODF3B, TYMP
rs2466029	chr8:127188726	■	■	■	■	
rs1002015	chr2:110859042	■	■	■	■	
rs210143	chr6:33579153	■	■	■	■	
rs142215530	chr15:56485493	■	■	■	■	
rs1036935	chr18:50317164	■	■	■	■	
rs41271473	chr1:228744549	■	■	■	■	CCSAP
rs6708784	chr2:111169802	■	■	■	■	BCL2L11
rs1317082	chr3:169779797	■	■	■	■	MYNN
rs3800461	chr6:34648545	■	■	■	■	C6orf106, SNRPC, TAF11
rs4869818	chr6:154150090	■	■	■	■	IPCEF1
rs6586163	chr10:88992261	■	■	■	■	ACTA2, FAS
rs2651823	chr11:2300420	■	■	■	■	C11orf21, TSPAN32
rs7254272	chr19:4069121	■	■	■	■	TBXA2R
rs57214277	chr4:184333619	■	■	■	■	
rs71597109	chr4:101819845	■	■	■	■	
rs9880772	chr3:27736288	■	■	■	■	
rs7705526	chr5:1285859	■	■	■	■	
rs9392504	chr6:412802	■	■	■	■	
rs73718779	chr6:2969044	■	■	■	■	
rs2267708	chr7:124752458	■	■	■	■	
rs4368253	chr18:59955055	■	■	■	■	
rs34004493	chr2:230289297	■	■	■	■	SP140, SP140L
rs73192661	chr3:188411006	■	■	■	■	LPP
rs77551289	chr18:63121512	■	■	■	■	VPS4B
rs874460	chr19:46673495	■	■	■	■	FKRP, PRKD2

Potential functional SNP (selected SNP)	Transcription factor	Risk allele	Binding affinity risk allele	Motif
rs210142 (rs210143)	SPH1	C	Decreased	
rs12591150 (rs142215530)	PAX5	T	Decreased	
rs4767033 (rs6489882)	NFE2A	T	Decreased	
rs319054 (rs11637565)	TCF3	G	Increased	
rs7701411 (rs4869818)	NFATC1	T	Increased	
rs3083367 (rs4368253)	FOXO1	G	Increased	

# Story 3: From 1D to 3D genomes in CLL

Huge impact on our understanding and the clinical management of hematological malignancies

Genetic changes

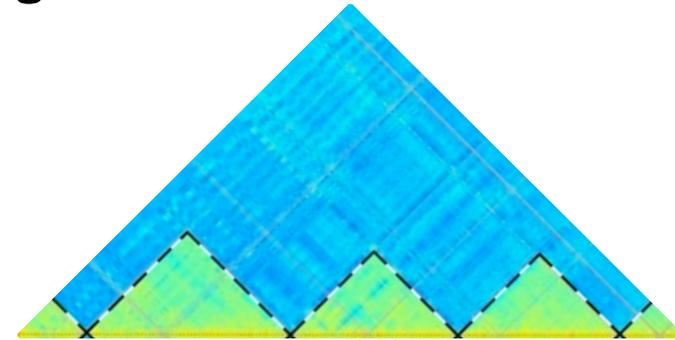
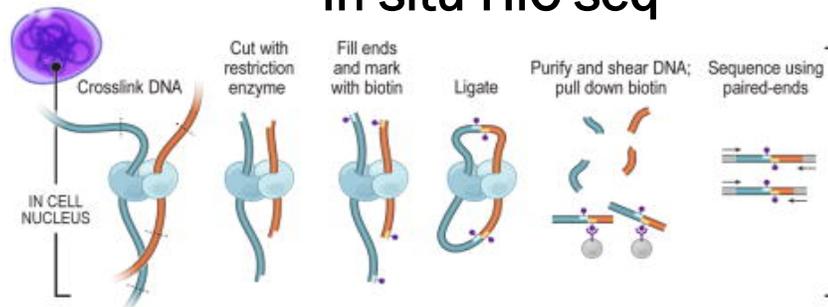
Gene expression changes

DNA methylation changes

Active (A) compartment

Inactive (B) compartment

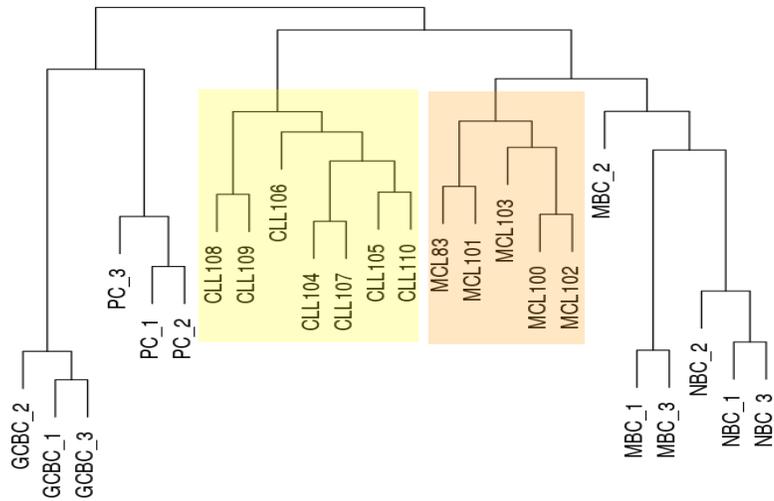
In situ HiC-seq



Active (A) compartment

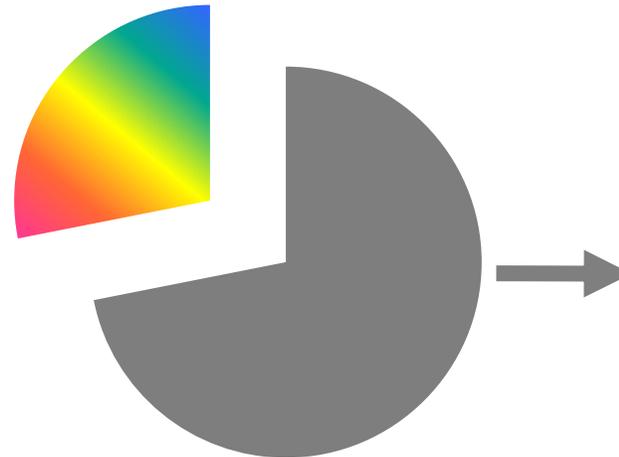
Inactive (B) compartment

# Story 3: Changes in the 3D genome structure



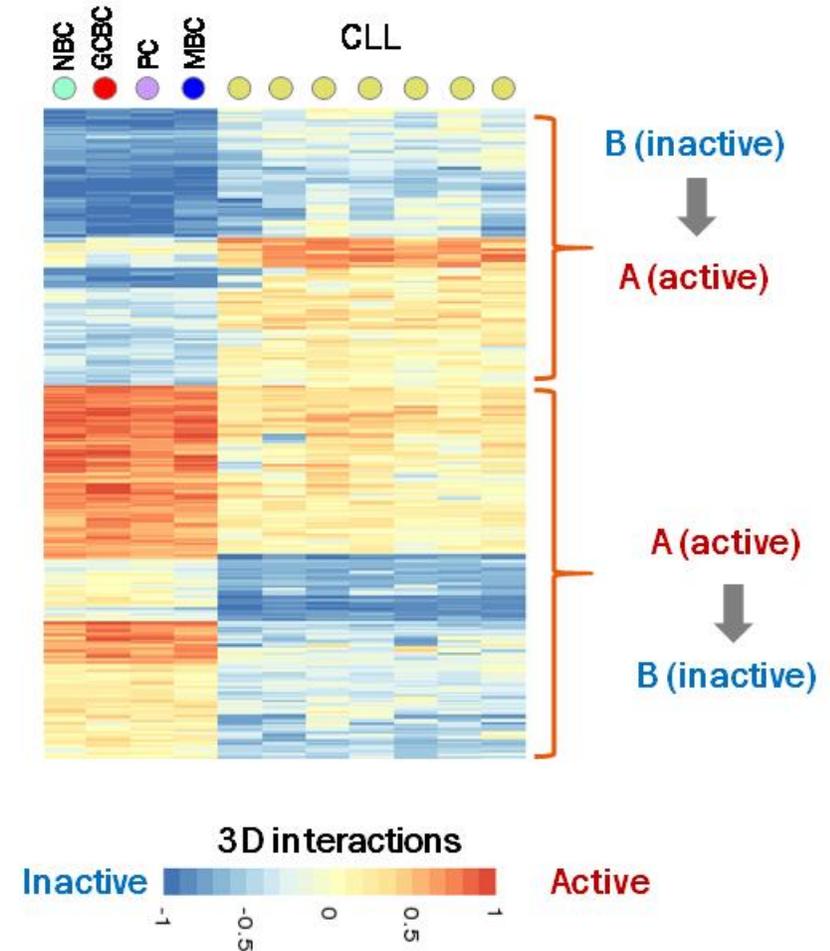
- The 3D genome structure changes during cell differentiation and is maturation stage-specific
- CLLs and MCLs cluster are in the same cluster as NBCs and MBCs
- CLLs and MCL show differential 3D structures

28% of the 3D genome changes during B-cell differentiation

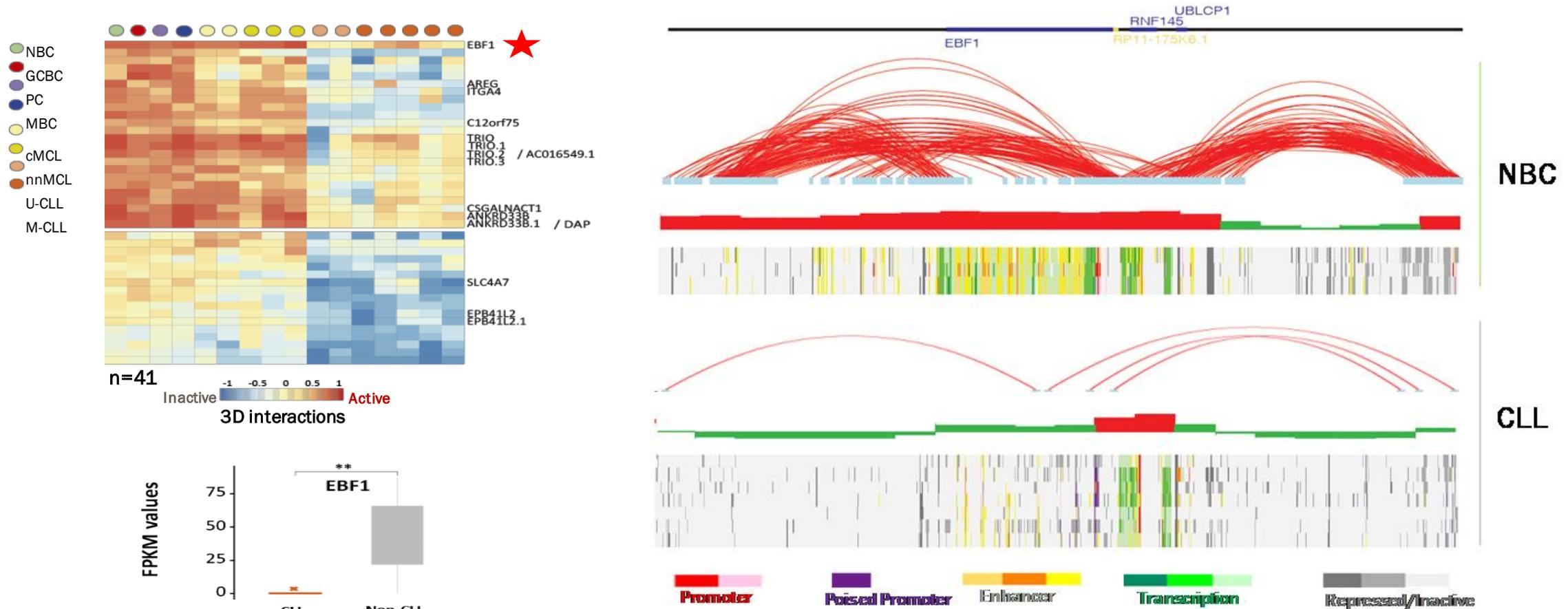


72% is stable in normal B cells

Can we detect changes in the 3D genome structure of CLL?

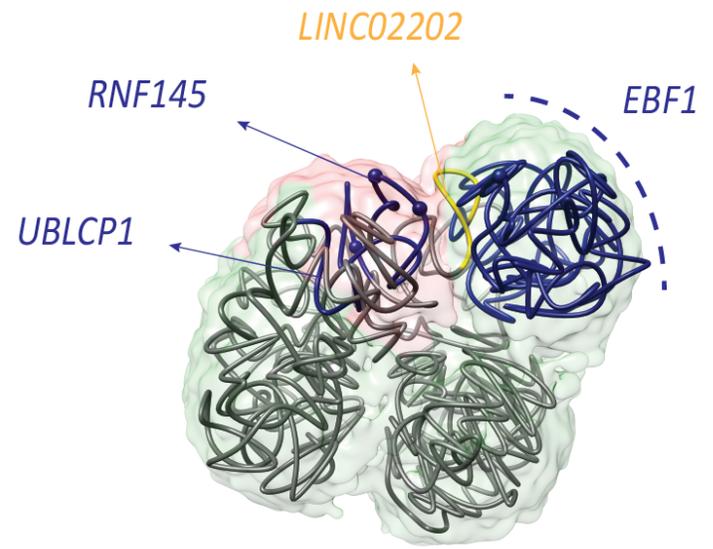
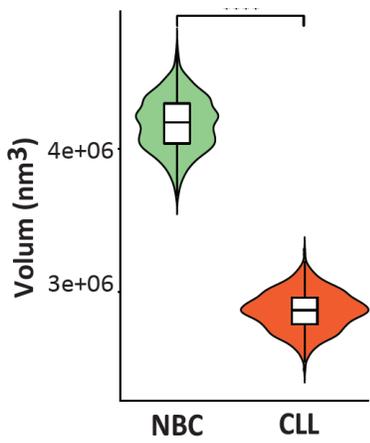
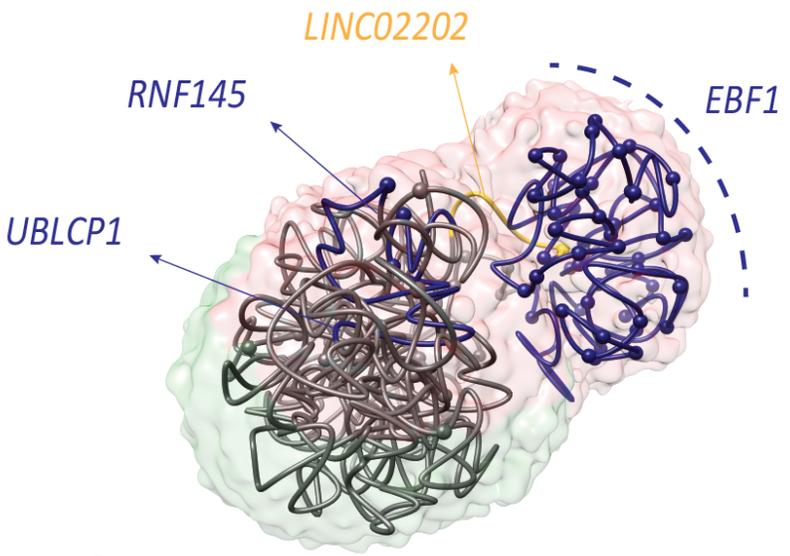
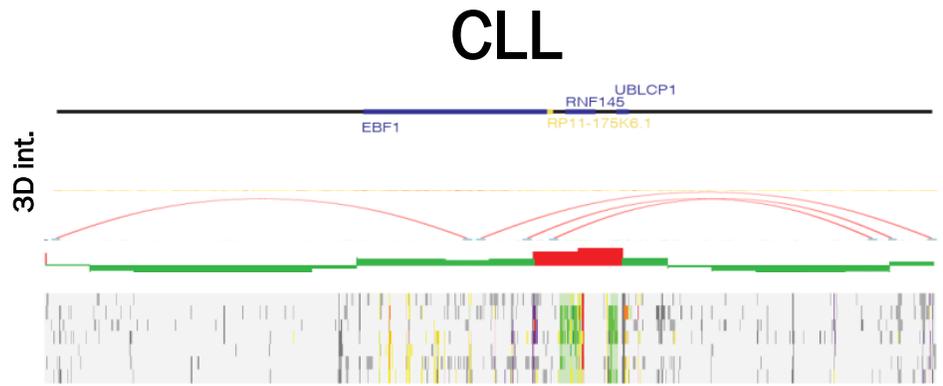
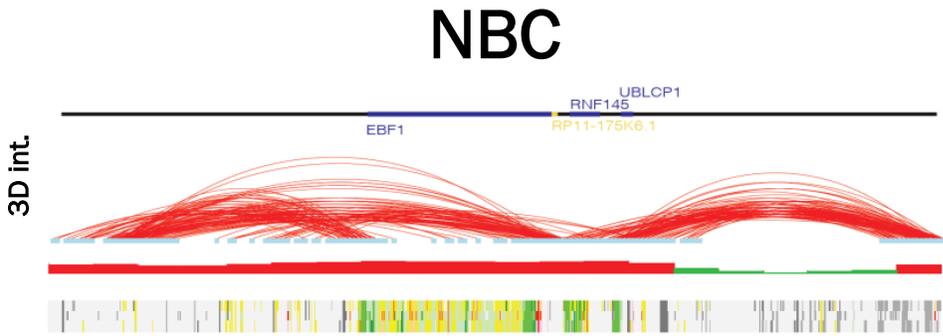


# Story 3: CLL-specific loss of 3D interactions



How is *EBF1* structurally organized into the nucleus?

# Story 3: Structural model of EBF1 in normal B cells and CLL



# *Take-home messages*

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- The regulatory chromatin landscape is extensively altered in CLL as a whole and in IGHV-based subtypes, and seems to be mediated by the action of few TF families.
- The great majority of the SNPs associated with CLL risk are located in regions enriched in regulatory elements.
- CLL shows an altered 3D genome structure and DNA blocks changing their level of 3D interactions contain genes related to disease pathogenesis.

# Acknowledgements

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